



# REAL MICROBIOME SOIL DNA KIT

Ref. RBMEGS18 50 PREPS

## 1. INTRODUCTION

The Real MICROBIOME Soil DNA kit has been designed for a fast and efficient purification of **microbial DNA from environmental samples like soil samples**

In this procedure, the microorganisms are efficiently lysed by a combination of heat, chemical and mechanical disruption with specialized beads. Inhibitors are eliminated by precipitation using a proprietary cleanup buffer. The sample is then applied to a microspin column and the DNA that is bound to the column undergoes a single wash step before elution.

### Features:

- **Designed for a fast and easy purification microbial DNA from different types of soil samples.**
- **Optimized lysis method-Combination of heat, chemical and mechanical lysis via bead-based homogenization enables isolation of DNA from yeast, fungi, Gram-negative and Gram-positive bacteria.**
- **Eliminates inhibitory substances as humic substances and others inhibitors.**
- **No phenol/chloroform extraction or ethanol precipitation is necessary.**

### Applications:

- **Microbiome analysis**
- **PCR applications.**
- **RFLP analysis.**
- **Patogen typing.**
- **Mutation analysis.**



## 2. COMPONENTS KIT

	<b>50 preps</b>	<b>Storage</b>
<b><u>Lysis Buffer Soil</u></b>	<b>30 ml</b>	Room temperature
<b><u>Enhancer Buffer</u></b>	<b>5 ml</b>	Room temperature
<b><u>EC Buffer</u></b>	<b>8 ml</b>	Room temperature
<b><u>Binding Buffer Soil</u></b>	<b>50 ml</b>	Room temperature
<b><u>Wash Buffer *</u></b>	<b>10 ml</b>	Room temperature
<b><u>Elution Buffer</u></b>	<b>10 ml</b>	Room temperature
<b><u>Bead Microtubrs</u></b>	<b>50 unidades</b>	Room temperature
<b><u>Collection Tubes</u></b>	<b>100 unidades</b>	Room temperature
<b><u>Microbial DNA Columns</u></b>	<b>50 unidades</b>	Room temperature

(\*) These solutions must be prepared as indicated in the **Preliminary Preparations** section of the protocol.

*PRECAUTIONS: The Lysis Buffer Soil and Desinhibition Buffer contains guanidium salts, and should be handled with care. Guanidinium salts form highly reactive compounds when combined bleach.*

### 2.1 Equipment and additional reagents required

- Microcentrifuge.
- Microcentrifuge tubes, DNase-free, 1.5 mL and 2.0 ml
- Ethanol 100%.
- Heat block, dry bath, or water bath (65°C).
- For vortex bead homogenization: hands-free adapter for vortex mixer, with horizontal tube orientation, we recommend the Vortex Genie 2.
- (Optional) alternative to vortex bead homogenization: Bead mill homogenizer.

## 3. PROTOCOL

### 3.1 Preliminary Preparations

- **Add 40 ml of Ethanol 100 %** to the Wash Buffer. Keep the container closed to avoid the ethanol evaporation.
- Pre-heat the Elution Buffer at 70°C.

### 3.2 General Remarks

#### **Sample Handly**

Collect samples according to your laboratory guidelines and experimental needs. Ensure that samples are mixed thoroughly with Lysis Buffer and Enhancer Buffer to create a homogenous sample. One way to ensure thorough mixing is to vortex the tube with the cap down.

#### **Sample Lysis**



The procedure is optimized by using "beads" in a vortex with horizontal agitation (Vortex Genie 2 or similar). Make sure that the vortex adapter allows horizontal agitation; This is a cost-effective method for recovery of high-quality microbial DNA.

Adapters with a vertical tube orientation may not agitate properly.

You can use "Bead mill" homogenizers such as FastPrep, Precellys and others but following the manufacturer's instructions to optimize the lysis of the sample. **IMPORTANT:** Many modern disruption devices can cause very high energy input in bead tubes. Depending on bead tube type and content (beads, liquid volume, sample type), especially high frequency of shaking and / or long shaking duration can cause breaking up of the bead tubes! **It is the responsibility of the user to perform initial stability test for the used bead tubes under the conditions used!** Perform initial test with water instead of lysis buffer and moderate machine setting (low frequency, short time) in order to avoid spillage of chaotropic lysis buffer in case of tube breakage.

### 3.3 Protocol for DNA extraction from 250 mg of environmental samples

1. Weigh **250 mg of soil sample** in a 2.0 mL **bead microtube** and add **600 µL of Lysis Buffer Soil**. Vortex.

Depending on the nature of the soil it may be necessary to increase the Lysis Buffer Soil and Enhancer Buffer proportionally with the aim of recovering 400 µL of supernatant in point 5.

2. Add **100 µL of Enhancer Buffer**. Mix well by shaking gently the microtube.
3. **Incubate at 65°C for 10 minutes.**
4. **Homogenize** by bead beating for **10 minutes** at maximum speed on the **Vortex Genie 2 or similar** using a **horizontal adapter**.
5. **Centrifuge at 14,000 rpm for 5 minutes.** Transfer up to **400 µL of the supernatant** to a clean microcentrifuge tube.  
**IMPORTANT:** A layer of debris may be present on top of the bead pellet. Avoid transfer of this debris with the supernatant.
6. Añadir **150 µl EC Buffer**. **Vortex**. Incubate at 0-4°C for 5 minutes.
7. **Centrifuge at 14,000 rpm for 3 minutes.** Transfer up to **500 µl of supernatant** in a new 1.5 ml microtube avoiding touching the pellet.
8. Add **900 µl of Binding Buffer Soil** and vortex briefly.
9. Load **700 µl** mixture sample into reservoir of a combined Microbial DNA Column-collection tube assembly. **Centrifuge at 8,000 rpm for 30 seconds.** Remove the collection tube.
10. Discard the flow-through, and repeat step 9 with the remaining sample mixture.

Ensure that the entire sample mixture has passed into the collection tube by inspecting the column. If sample remains in the column, centrifuge again at 14,000 rpm for 1 minute.

11. Add **700 µl of Wash Buffer**. **Centrifuge at 14.000 rpm for 1 minute**. Discard the flow-through.

**12. Dry silica membrane.** Centrifuge at 14.000 rpm for 3 minutes.

13. Place the Microbial DNA Column into a 1.5 mL nuclease-free tube (not provided) and add **50-100 µL Pre-heat the Elution Buffer** at 70°C. Incubate **at room temperature** for **2 minutes**.

**14. Centrifuge** the spin column-tube assembly **at 14.000 rpm for 1 minute**, then discard the column. The purified DNA is in the tube.

#### **4. PROBLEM GUIDE AND POSSIBLE ANSWER**

Due to the great environmental samples variety that can be treated, it becomes difficult to generalize possible problems and answers. For this reason, we recommend to contact **Durviz** Laboratory Technical Service for any question regarding the protocols, specific soil samples or any problem you may have during the process.  
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