NUEVO Ed. 01-2019

REAL MINISYSTEM + CONE+ STICKS

Product for *in vitro diagnostic*.
150 TESTS RPP9000MD

COMPONENTS:

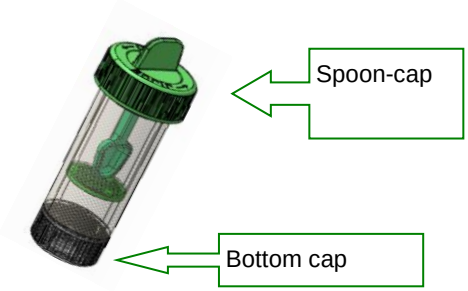
- Sedimentation cone (ref. CS01).
- Sticks for sediment collection (ref. PR01).

INDICATIONS:

- Avoid the use of antidiarrheal or laxative drugs prior to sample collection.
- To ensure the recovery of parasites it is recommended to collect three samples of the patient on several successive days for examination.
- Wash hands with soap.
- Use before the date indicated on the package.
- Store at room temperature.

SAMPLE COLLECTION

1. Defecate in a clean, dry container.
2. Prevent feces from being contaminated with urine or water.
3. Choose an appropriate amount of stool, collect with the spoon-cap of the tube and insert in the middle (a full tablespoon). The amount of sample to be collected should not exceed the capacity of the tube. Do not open the lower tube plug at any time.
4. Close the tube tightly and send it to the laboratory for processing



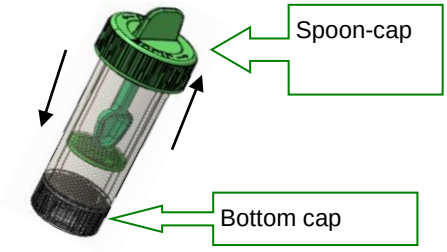
DIAGNOSTIC METHODS

A. Direct Microscopic Examination

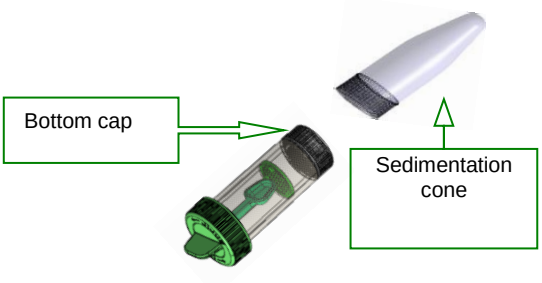
1. Prepare the smear by mixing a small amount of preserved fecal material (approximately 2mg) with one drop of physiological saline solution in a glass slide.
2. Cover slides with a coverslip (22 x 22mm).
3. Examine the entire coverslip using the low magnification target (X100, 10X magnification). If something suspicious is observed, increase the target (X400, 40X magnification). Through direct microscopic examination eggs and / or larvae of helminths and some cysts of protozoa can be observed. Since the fecal material has been preserved, the organisms will have no mobility.

B. Concentration technique

1. Shake the collection tube vigorously with the sample for 30 seconds.
NOTE: The sample should remain in the middle at least for 30 minutes.



2. Optionally, if the sample has a very compact appearance, remove the cap from the tube and add 3 drops of surfactant (Triton X-100 to 20%) to improve its homogenization.
3. Mix the tube contents by shaking vigorously for 30 seconds.
4. Optionally, if the sample has many residues, add 1 ml of ethyl acetate and stir for 30 seconds.
5. Replace the lower plug of the tube by the settling cone.



Durviz s.l. C/Leonardo da Vinci, 10 Parque Tecnológico de Valencia, Paterna - Valencia – Spain
Phone : +34 96 136 61 07, fax +34 96 136 61 68 Email : durviz@durviz.com – Website : www.durviz.com

6. Centrifuge at 700 xg (2000rpm) for 3 minutes.

7. Collect sediment by using a pasteur pipette. Remove the cone with the sediment and decant the supernatant. If a very thick sediment is observed, it can be resuspended with saline solution (2-3 drops). Place a few drops on a microscope slide and observe the presence of intestinal parasites (Giardia intestinalis, Entamoeba histolytica, Blastocystis, hominis ...).

c. Permanent Satining

If you want to do some staining, prepare a slide and add a small amount of sediment, spread the sediment on the slide to prepare a fine smear.
Allow to dry overnight at room temperature or for several hours (from 30 minutes to 60 minutes if the slide is thicker) in an incubator at 37 ° C.
Do not use heaters, these can affect the organisms present in the sample.
NOTE: Since Total-Fix® does not contain PVA (which helps the smear to stick to the slide), albumin may be added to the slide before preparing the fecal smear. However, if the smear is completely dry before staining, it is not necessary to add albumin.
Finally, proceed with the staining of choice. Trichrome staining is recommended, although iron hematoxylin may also be used.

PRECAUTIONS:

- Avoid contact of the Total-Fix® solution with the skin or eyes. In case of contact, wash affected area with water.
- If irritation develops, contact a physician immediately.
- The Total-Fix® solution is toxic. If accidental ingestion occurs, drink milk or water and contact a physician immediately.
- Each sample should be treated as a potential source of infection.
- Correct laboratory guidelines should be followed at all times.
- It is recommended to wear gloves and wash hands.

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